

Scaling for ATMP commercialisation

Cell and Gene Therapy Catapult

Daria Marsh, Head of Bioprocessing

We are an independent innovation and technology organisation committed to the advancement of the cell and gene therapy industry



Our vision

A thriving industry delivering life-changing advanced therapies to the world.



Our role

Create powerful collaborations which overcome the challenges to the advancement of the ATMP sector.



How we work

We are a team of experts covering all aspects of advanced therapies. Applying our own unique capabilities and assets, we collaborate with academia, industry and healthcare providers to develop new technology and innovation.

ATMP Demand and the Patient Access Challenge

- Patient demand for commercially approved ATMP products outweighs current supply
- CAR-T pricing >\$180k; ~\$1M with cost of administration. Drive to improve margins to ~80% by 2030.*²
- In the Allogeneic space at least 1,200 patients presenting with 34 different indications have been implanted with an accumulated dose of at least 190 billion hPSC derived cells + 200 billion platelets.*⁴

*¹ Patient Access Analytics | Ori Biotech, OriBiotech (2024)

*² Alliance for Regenerative Medicine Meeting at the Med 2025

*³ Driving the next wave of innovation in CAR-T cell therapies: McKinsey (2019)

*⁴ Kirkeby et al., 2025, Pluripotent stem-cell-derived therapies in clinical trial: A 2025 update, Cell Stem Cell 32, January 2

*⁵ Adapted from ARM Cell & Gene State of the Industry Briefing, EvaluatePharma, Deloitte Analysis, Jan 2025

CAR-T Products

~10,000

CAR-T products
manufactured/year*¹

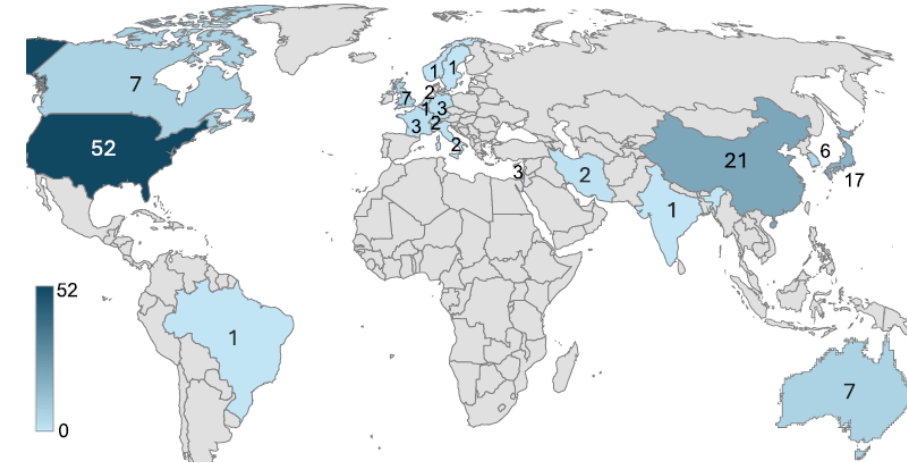
~1,894

Clinical trials*³
underway

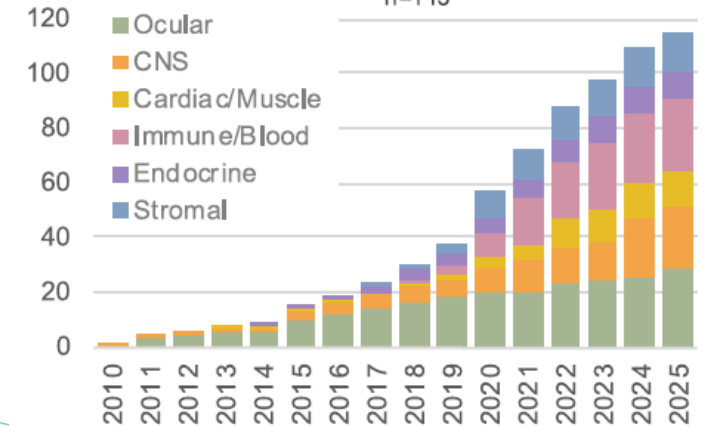
2M

Hematological
cancer patients
eligible for CAR-T
by 2029*³

hPSC Derived Products*⁴



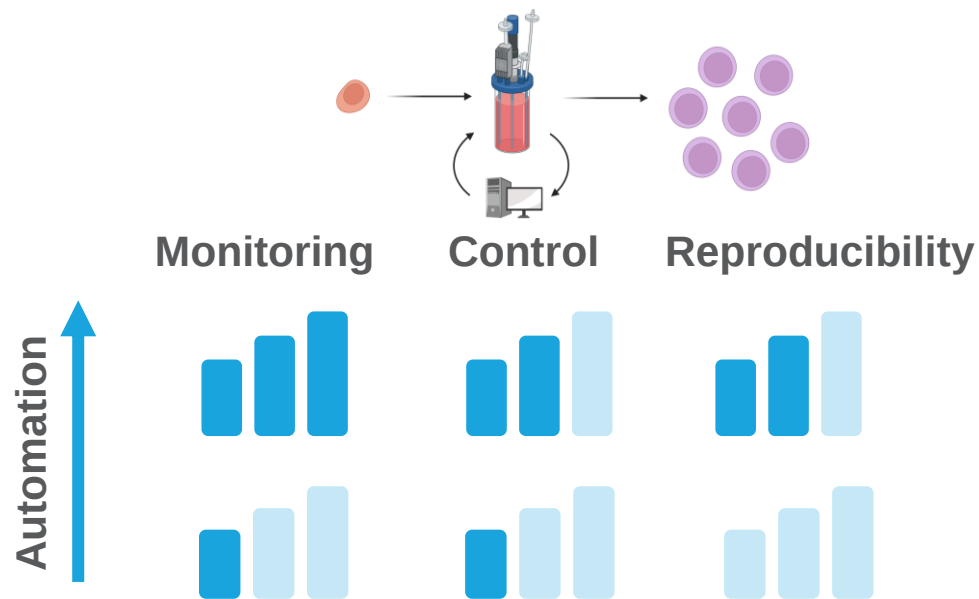
Cumulative trials over time
n=115



Challenges for Scaling

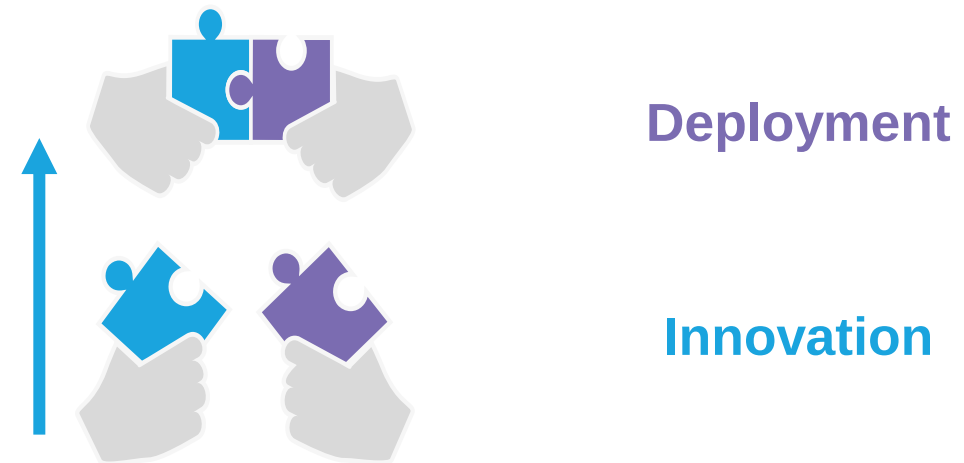
Scale up: iPSC derived case example

1. Close the process
2. Maximise yield and quality
3. Use PAT to automate control



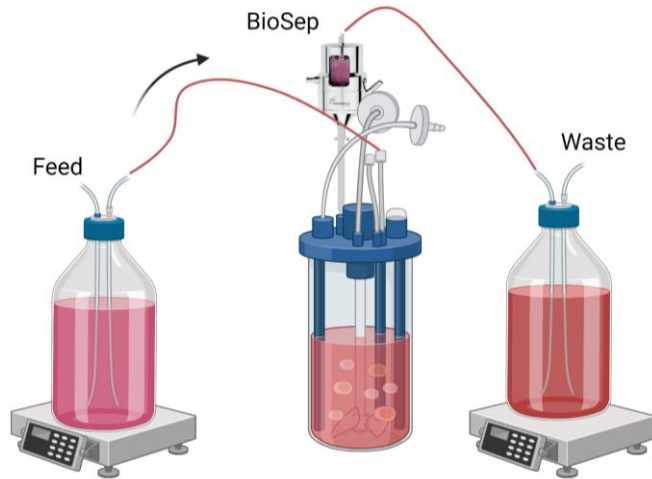
Scale out approach

1. Min intervention, max reproducibility
2. Automate for controlled parallelization
3. QC Automation
4. Digital Integration & Advanced Control

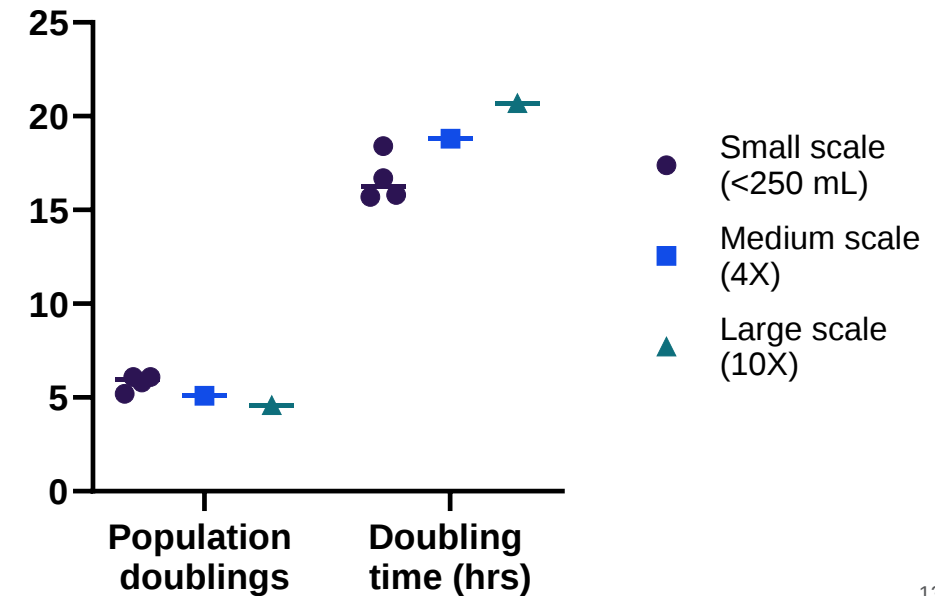
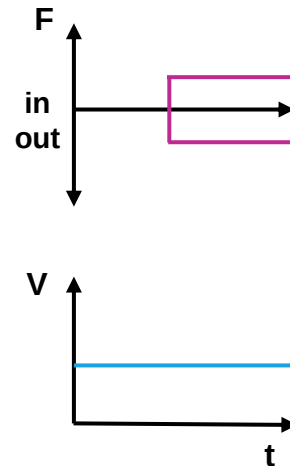
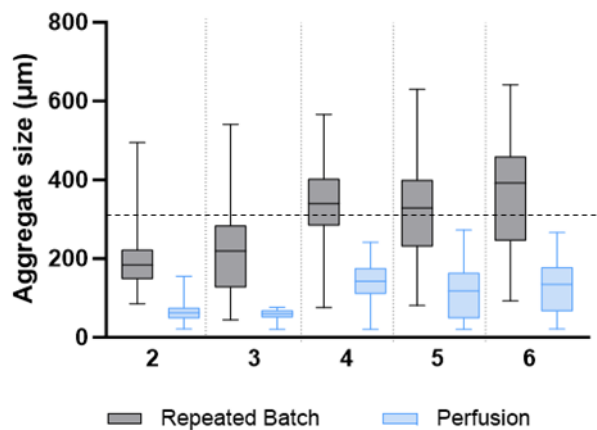


Scale Up: iPSC Expansion

Close process, Maximise Yield and Quality

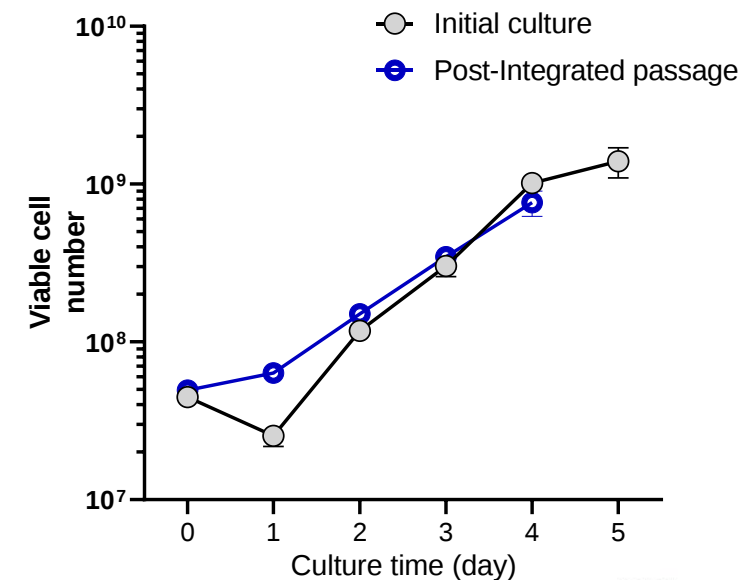
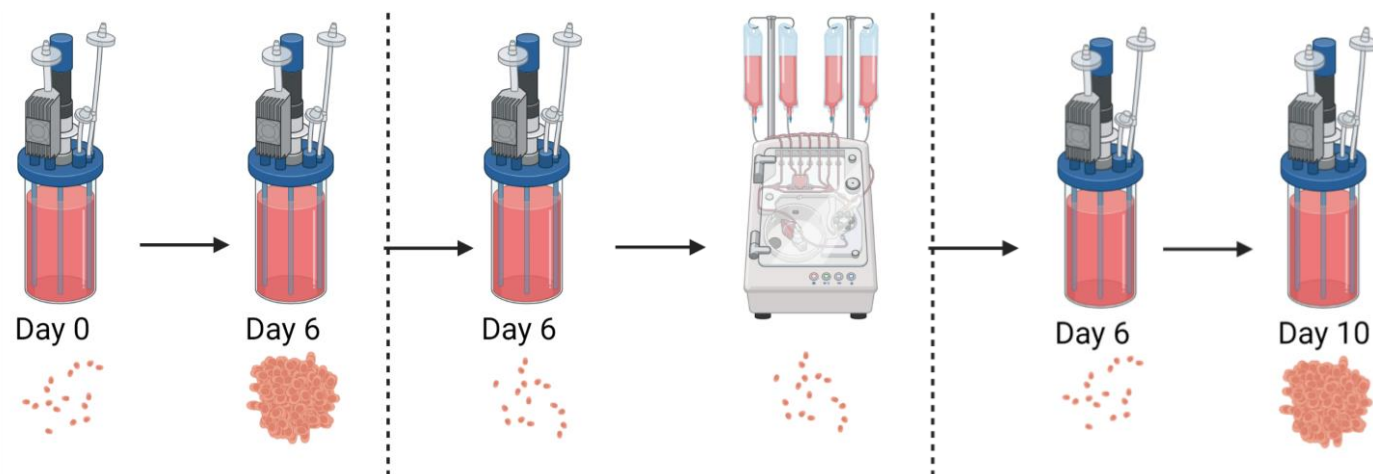


- Intensification: perfusion controlled cell expansion process in STR
- 30-fold expansion of iPSC in 5 days (**10^7 cells/mL**)
- More homogenous aggregate size
- Potential for further intensification and seamless downstream differentiation



Scale Up: iPSC Integrated Passage

Close process, Maximise Yield and Quality



Semi-Automated integrated passage to facilitate scale up



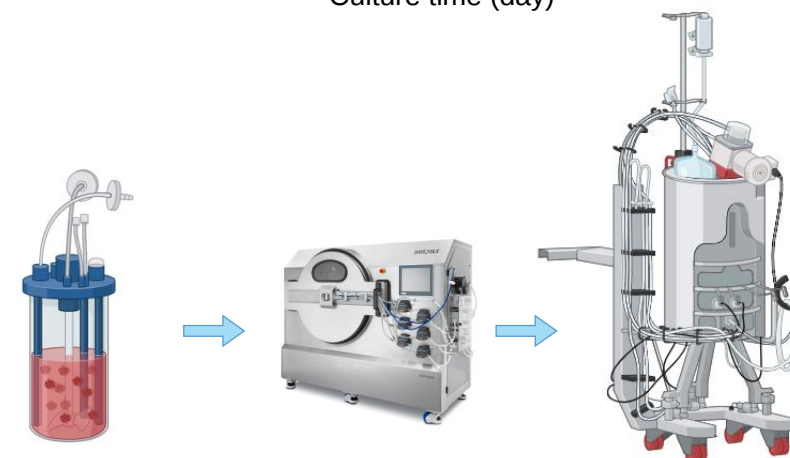
Retains **closed processing** for all intermediate steps



Proven at <250 mL scale



>80% cell recovery achieved for in situ dissociation and inoculation into subsequent culture



PAT for Aggregate Size

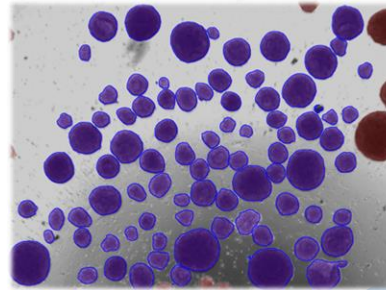
Aggregate size is a key in-process measurement

- Linked to PSC quality and differentiation efficiency

Existing manual workflows are inefficient with poor data integrity

- Manual
- Several points of manual data transfer and analysis
- Out-dated image analysis pipeline

Multi-day to 15 min workflow

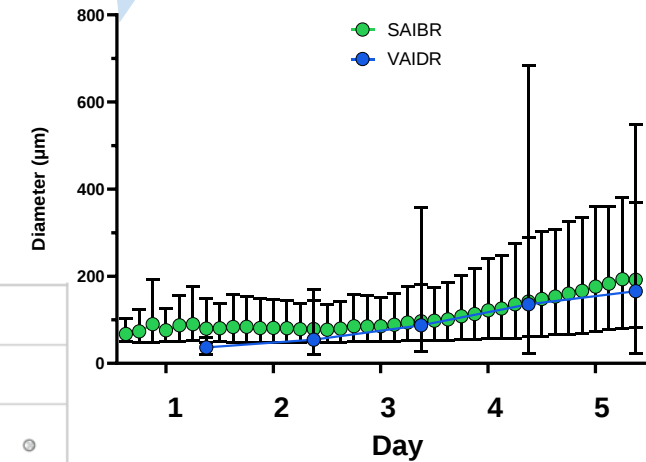
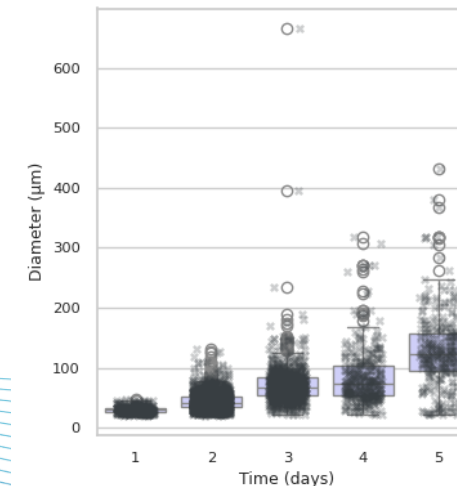
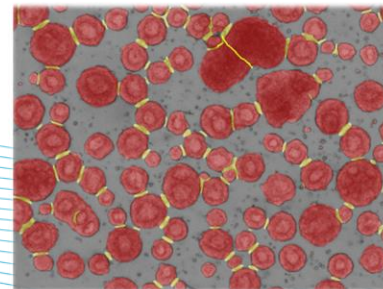


Train

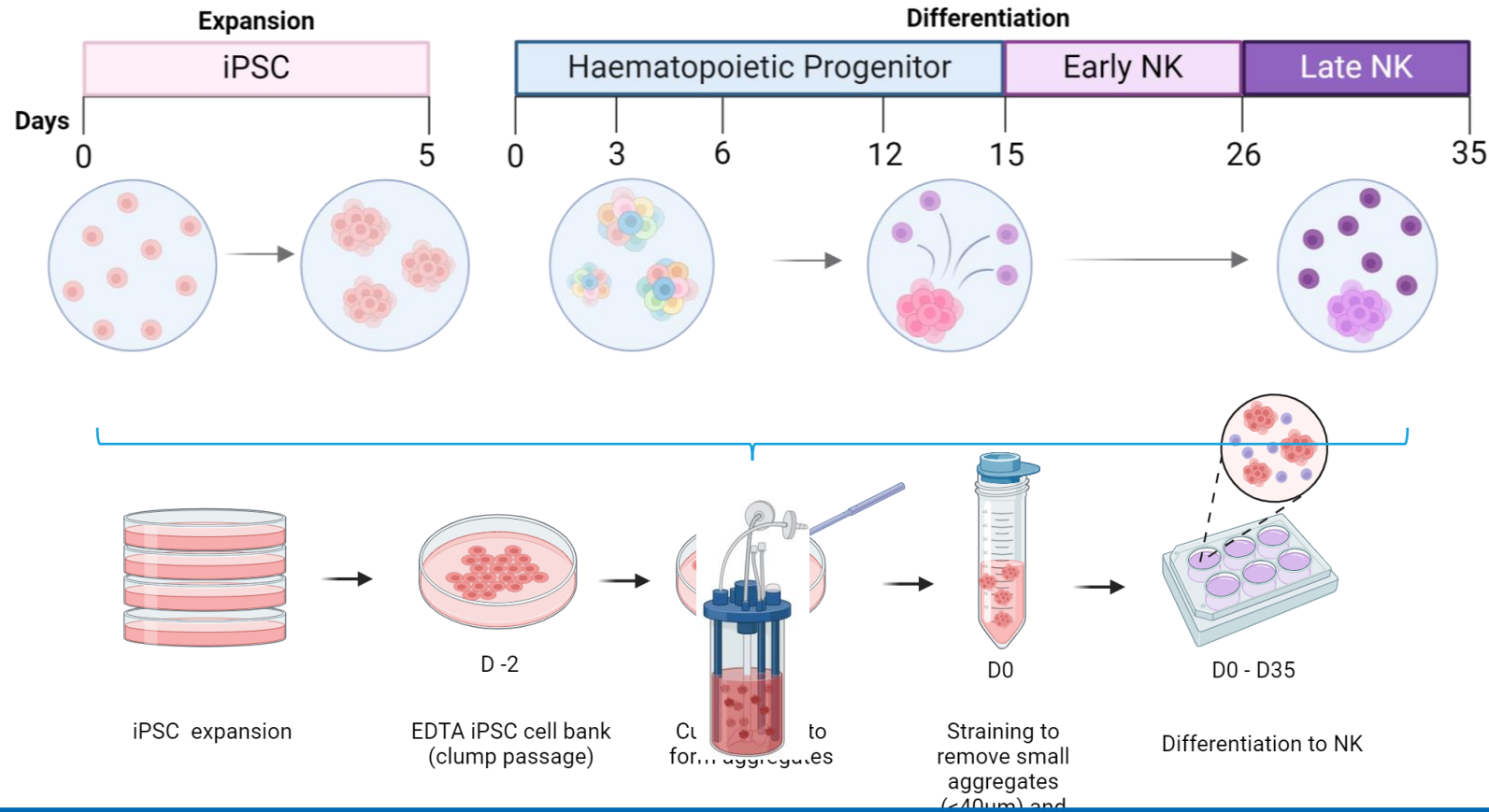
Analyse

Report

Integrate



Scale Up: NK Differentiation Case Study



Integrated iPSC expansion and NK differentiation in an STR system

Closed and Maximised Yield and Quality of iNK

YIELD

Integrated iPSC to NK differentiation in an STR, resulting in functional NK cells

- 10X improvement in yield
- High purity of NK cells

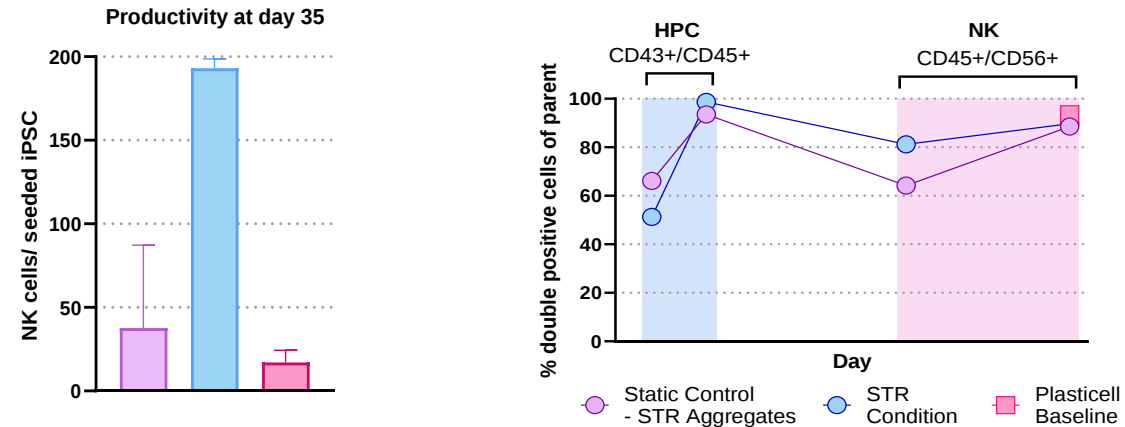
QUALITY

STR-generated iNKs showed cytotoxic function against 3 cancer cell lines.

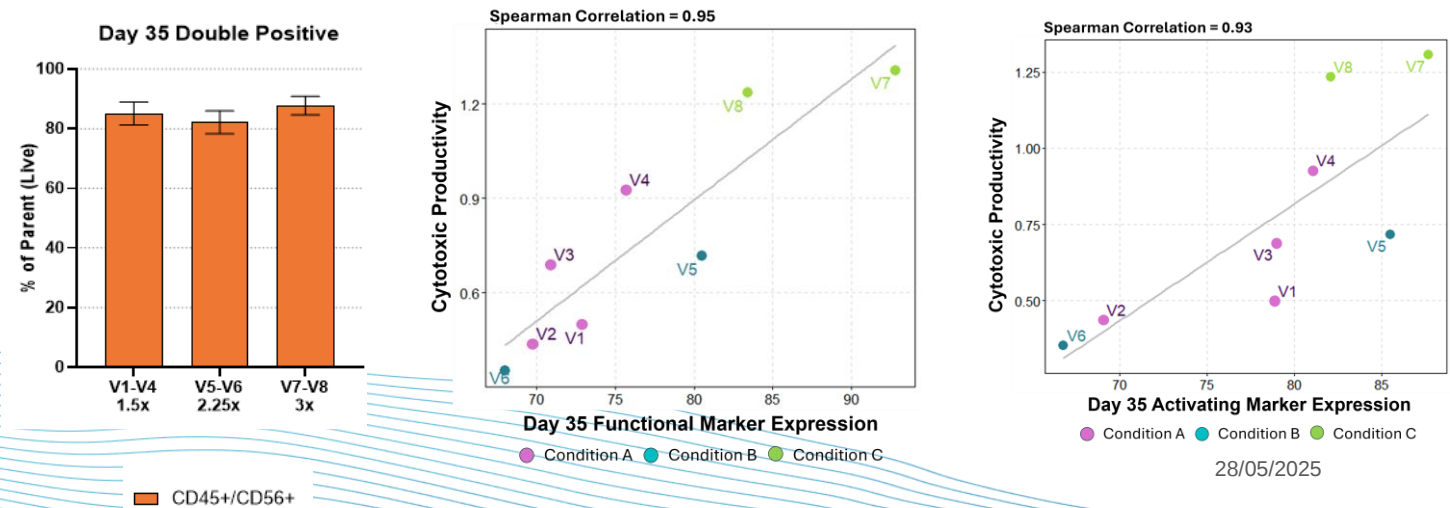
Correlating these outcomes with FCM data, revealed different phenotypes associated with:

- iNKs from processes that were highly productive.
- iNKs that were highly functional.

Feasibility Assessment



STR Condition Screening



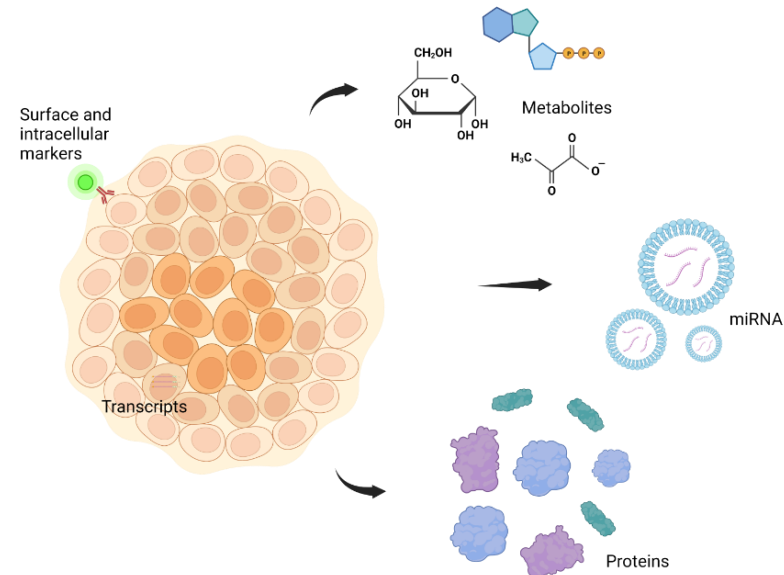
Product Characterisation

scRNAseq Screening

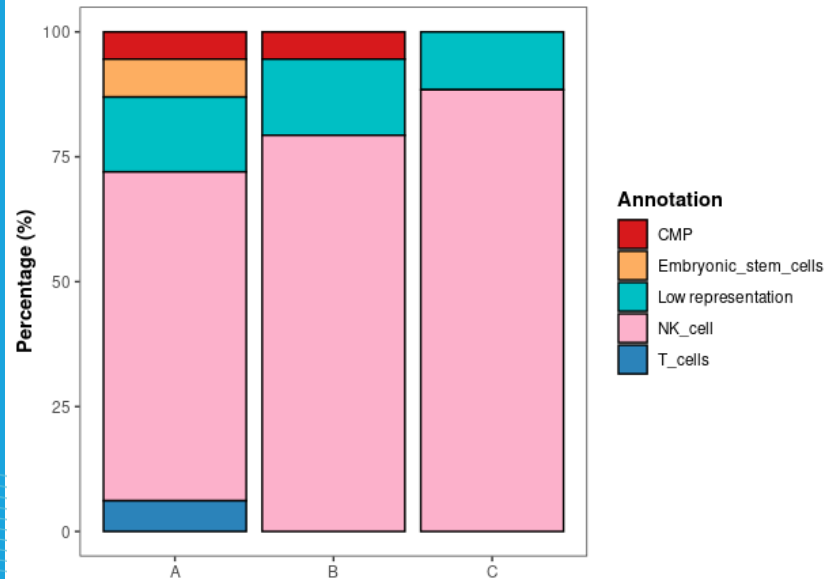
- Louvain clustering at day 35 - presence of different cell populations per condition, unique clusters in condition C.
- Modulation of cell density at a key transitional timepoint influenced the proportion of differentiated cell populations
- Cluster annotation (Human Cell Atlas) showed the largest population in all STR conditions consisted of target iNK cells, with small proportions of off-target cells

Targeted Protein Screening

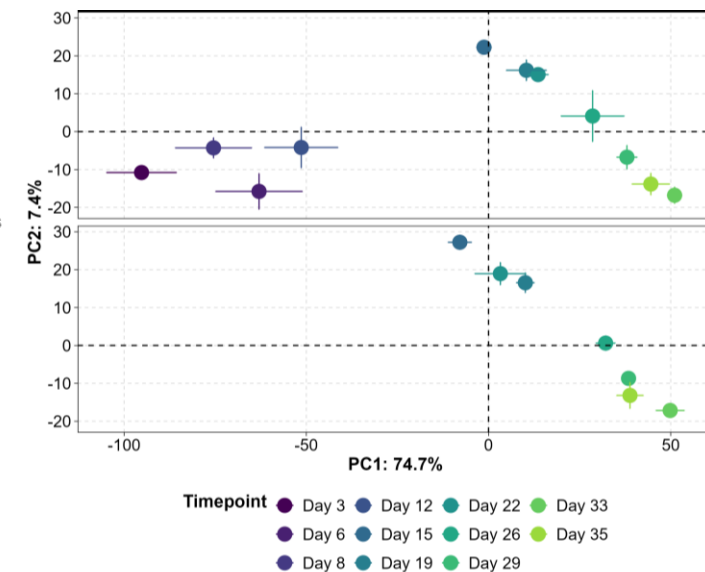
- Olink screening platform
 - Uses proximity extension assay for high specificity
 - High throughput screening of 5400 proteins
 - PCA of 1010 proteins
- Selected proteins capture shift in proteome during NK differentiation



scRNAseq



Targeted Protein Screening



Gain greater understanding and control of your product and process through use of **in-line analytics and real-time data interpretation** to adjust and improve process performance



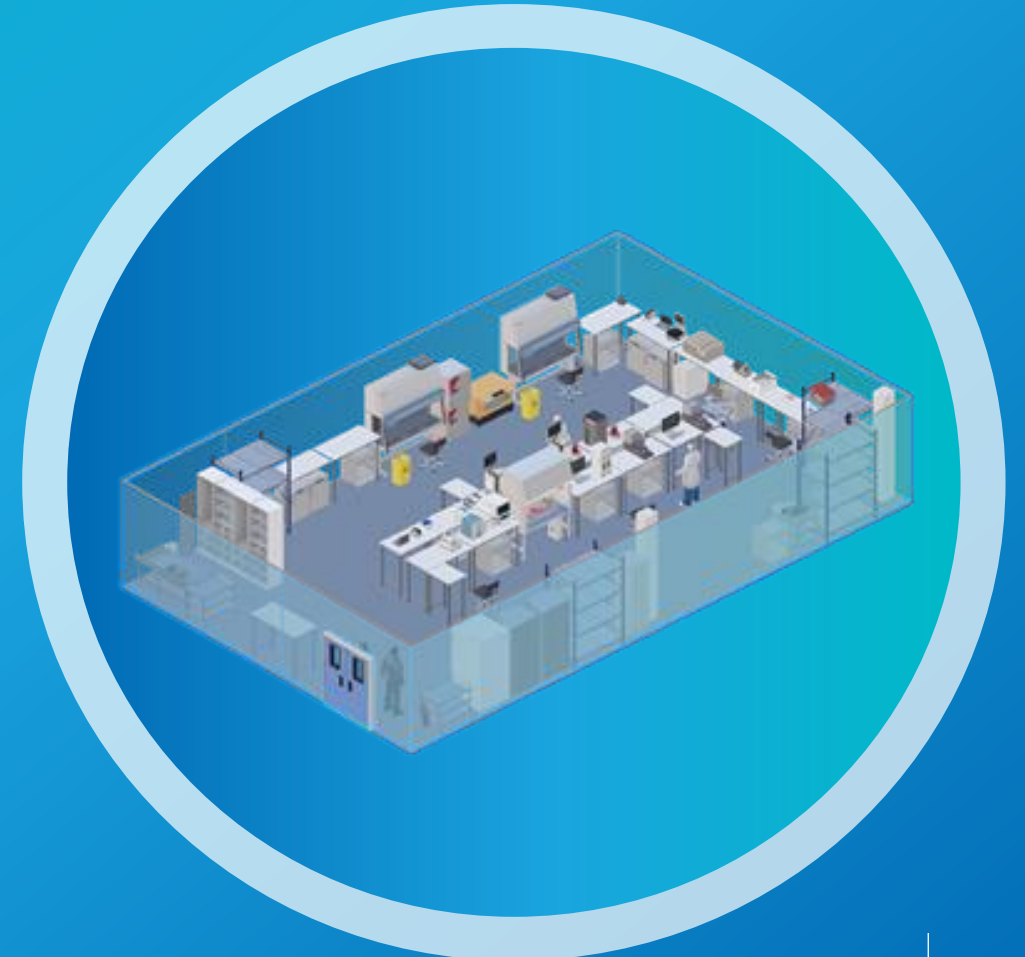
State-of-the-art **lab automation and digitalisation** allow rapid, high-throughput sample collection and analysis



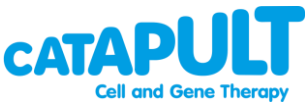
Assess and integrate **process analytical technologies** and advanced analytical tools to build in-depth characterisation



Develop **digital twins and advanced process control strategies** to maintain critical process parameters and critical quality attributes



Digital Twin Build



Digitalisation and Automation of
Medicines R&D and Manufacture

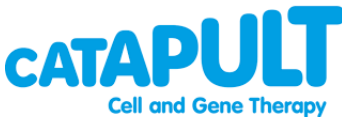
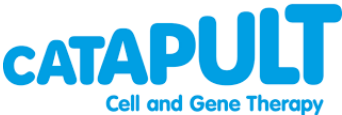
SMARTER Consortium

Advanced Control Development
for AAV Manufacture

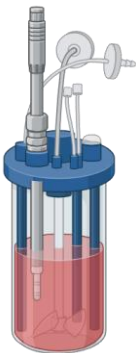
Advanced Control Development
for TIL- like cells and DCs



Leibniz
Universität
Hannover

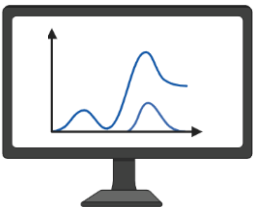


Spectroscopy probe
(real-time data-analysis)



*Soft sensor
prediction of CPP*

Model-predictive
advanced control

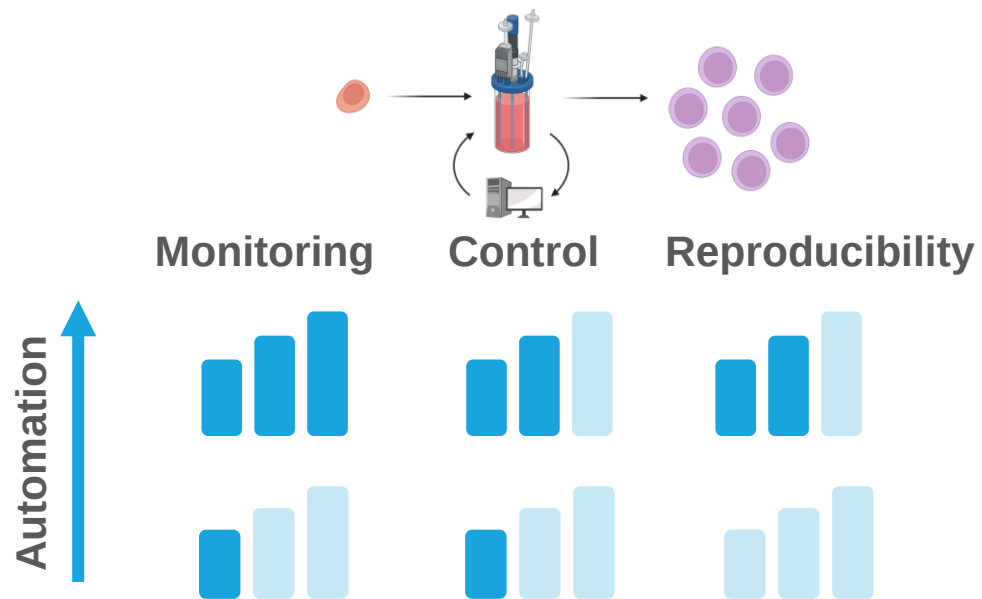


*Real-time process
adjustment*

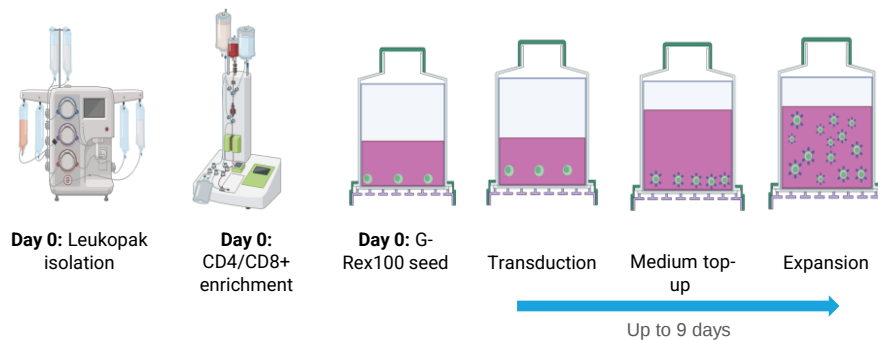
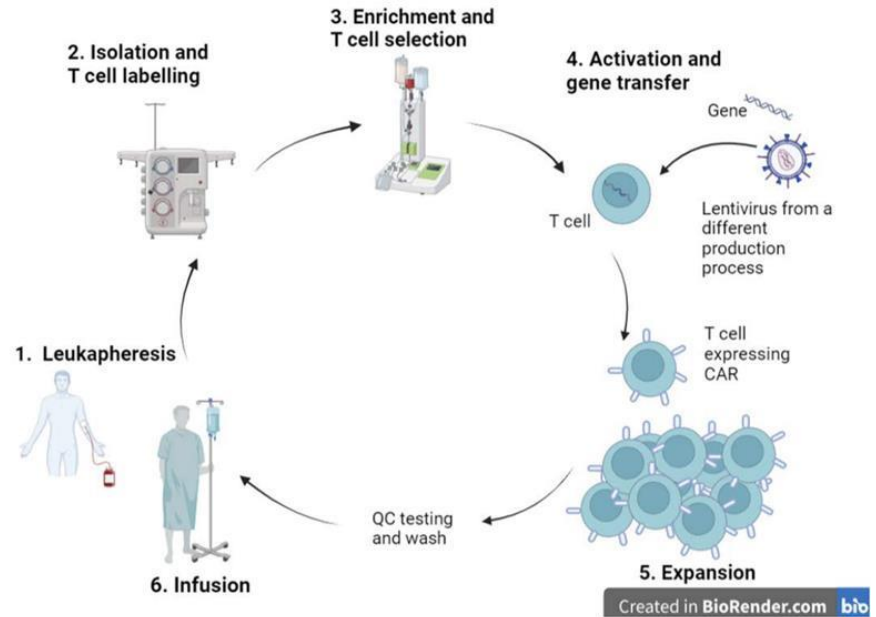
Challenges for Scaling

Scale up: iPSC derived case example

1. Close the process
2. Maximise yield and quality
3. Use PAT to automate control

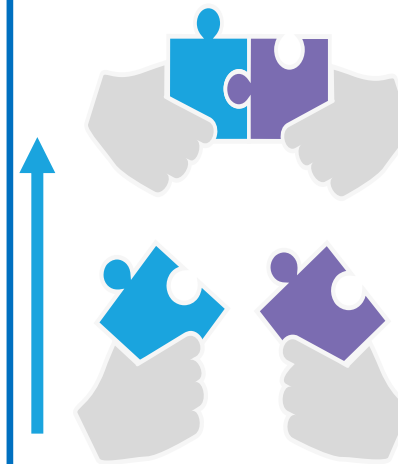


Challenges for Scaling



Scale Out: CAR-T case example

1. Min intervention, max reproducibility
2. Automate for controlled parallelization
3. QC Automation
4. Digital Integration & Advanced Control



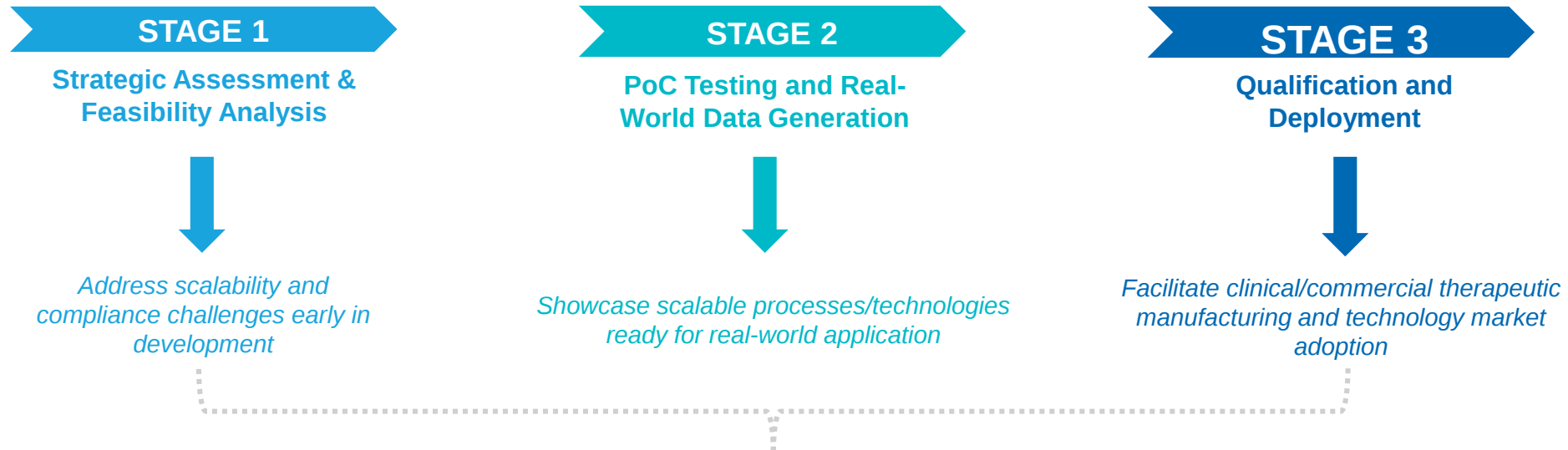
Deployment

Innovation

Automate for controlled parallelization

Expert guidance and tailored activities in a controlled environment for de-risked testing, validation and adoption of next-generation technologies.

Phased Journey to Power Scalable Innovations



De-risk and accelerate time-to-market and strengthen value proposition



QC Automation: challenges of process scale up in manufacturing

25%* cost per batch attributed to QC/QA

QC bottle necks

- Reliance on multiple offline instruments
- Inherent variability of living samples – SMEs required
- Labour intensive manual release testing
- Lack of standardisation of methods/instruments

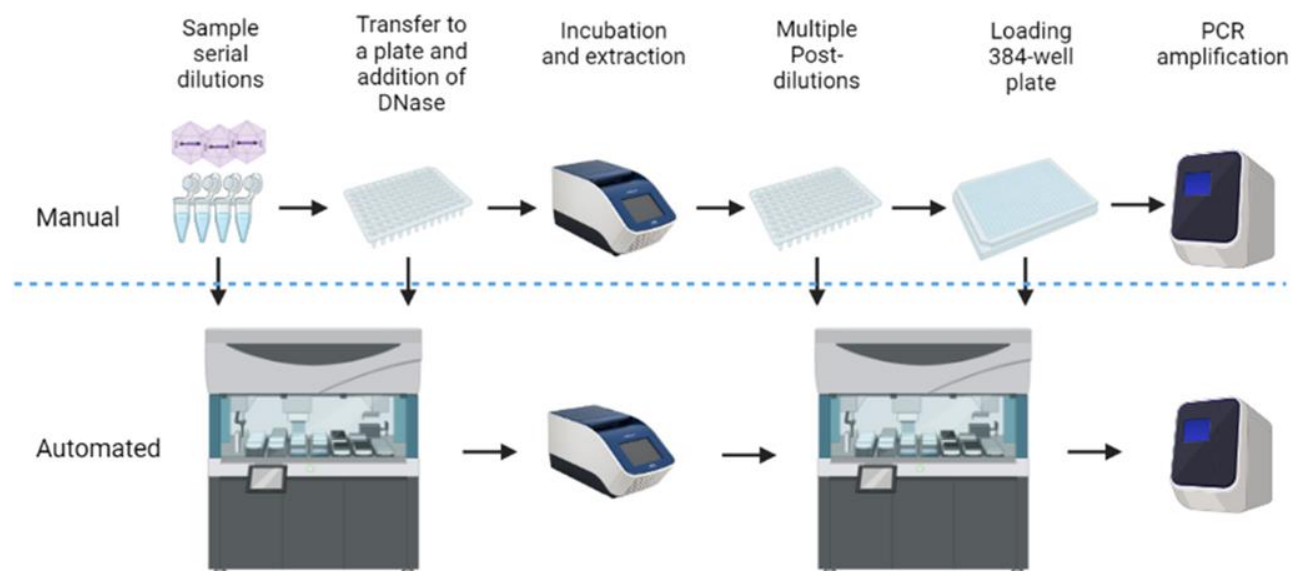
Need for automation/digitalisation

- Increase throughput – fast release e.g. autologous cell therapy
- Improve quality – reduce variability, improve data integrity and reduce error
- Reduce labour intensive analytics
- Reduce overall complexity



Case Study – Introducing assay automation

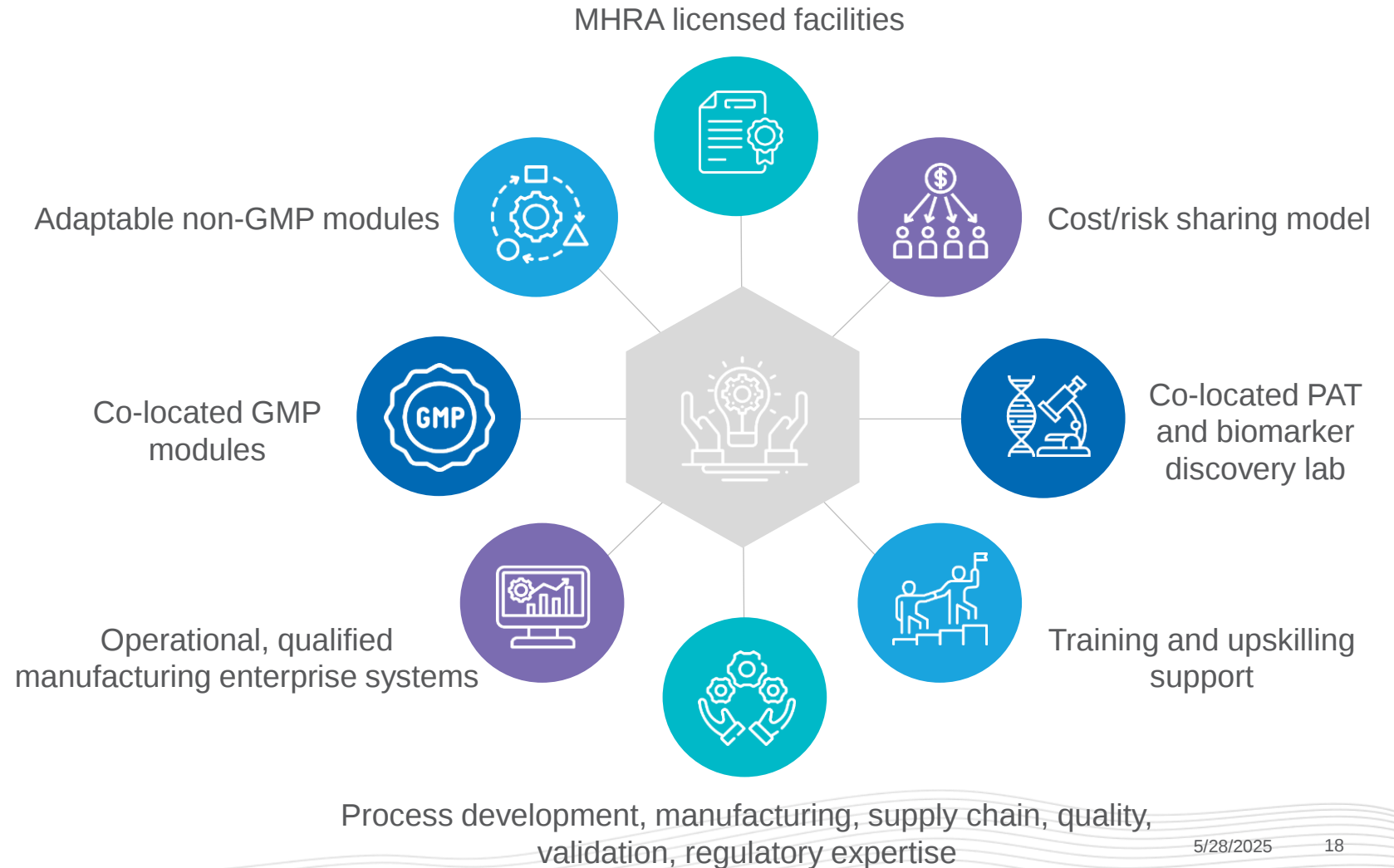
Improvements in throughput, quality, reliability and data integrity



Workflow	Manual	Automated
Lab Hands-on time	6h/plate	2.5h/plate
Samples per run	Up to 15 samples	Up to 32 samples
Sample pass rate (%)	36%	96%
Costs per sample (%)	100%	56%

Digital and Automation Testbeds

“First-in-the-world” ATMP-specific **GMP-mirroring** testbeds for **de-risked**, **accelerated** testing and adoption of next-generation **automation** and **digital technologies**.

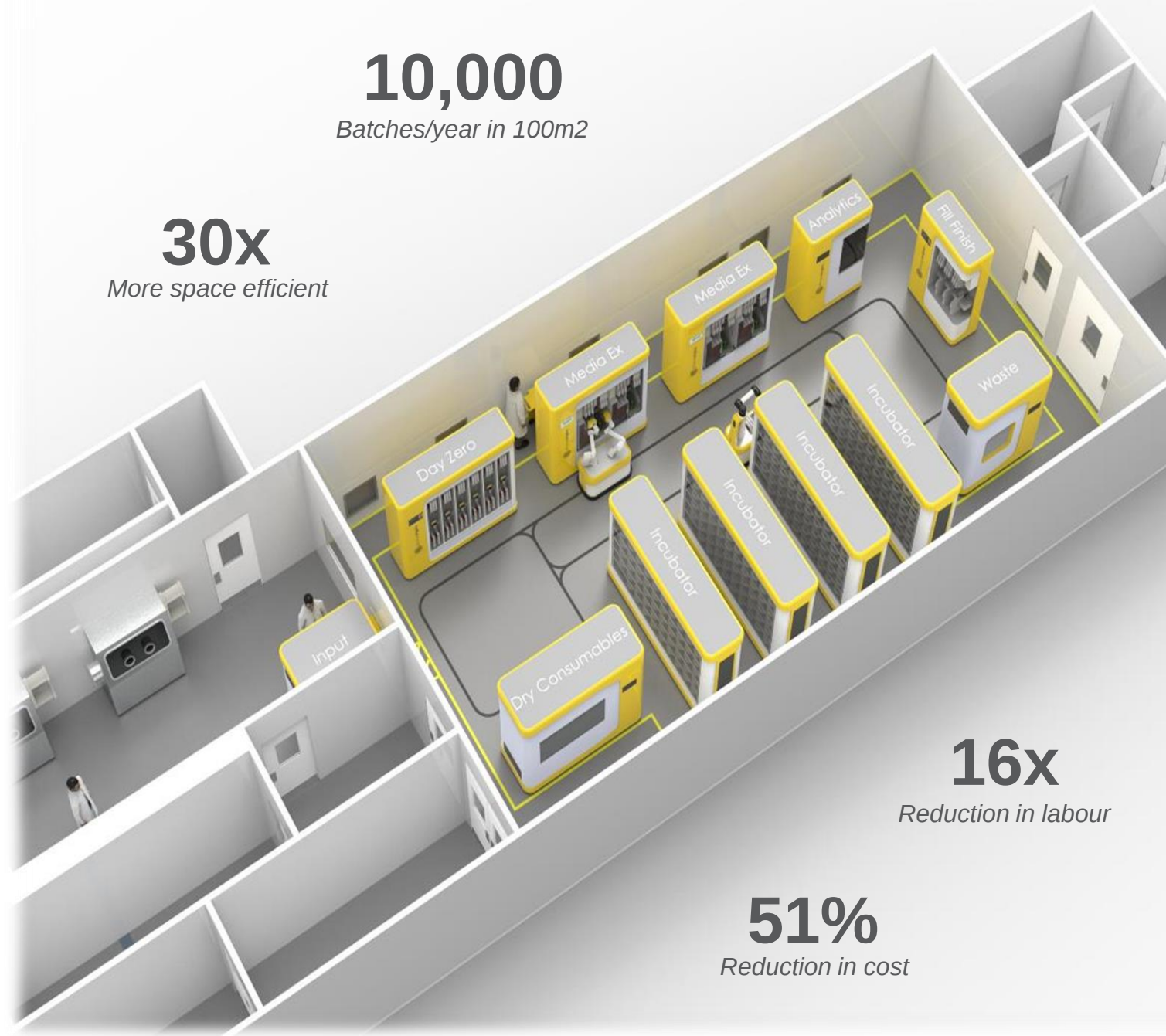


Future Factory Concept

Integration of fully automated robotics and PATs for **scalable**, **reproducible**, **high-quality** and more **environmentally sustainable** manufacturing of CGTs.

Ensures regulatory **compliance**, product **consistency** and a **scalable** route-to-market.

Increased attractiveness of innovations for investors, enhancing the **exit potential** and overall **commercial value**.



Scalable Therapy Development

CATAPULT
Cell and Gene Therapy



Skilled cell biologists, analytical scientists, bioprocess engineers, bioinformaticians



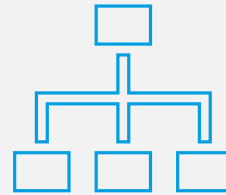
Developing robust, scalable manufacturing processes for pluripotent stem cell production



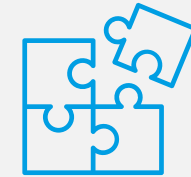
Knowledge of high-content metabolomics, and transcriptomics analysis to enable PSC analytics development



20+ collaborators supported



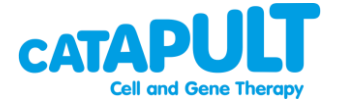
Broad experience across cell therapy modalities



Disruptive innovation through key collaborative projects



Thank you to everyone involved



CGT Catapult

Dr Jahid Hassan
Aishwarya Nair
Vera Karels
Robert Podovei
Antonia Karachaliou
Dr Patrick Statham
Dr Charlotte Lee-Reeves
Mudith Jayawardena
Dr Simona Zingaro
Dr Marianne Henry
Dr Virginia del Solar Fernandez
Juline Guenat
Dr Marcia Mata
Dr Chris O'Grady
Dr Dragos Marginean
Dr Joseph Oddy

Coralie Nerincx

Dr John Churchwell
Dr Molly Tregidgo
Kevin Vela
Lee Elkington
Rhys Macown
Mohsen Shaeri
Andrew Riley
Majahar Sayed
Su Jayakody
Julie Kerby
Dr Stephen Ward

Plasticell

Dr Marina Tarunina
Dr Tanya Ponomaryov
Sachin Luharia
Matthew Houppermans
Lam Lam

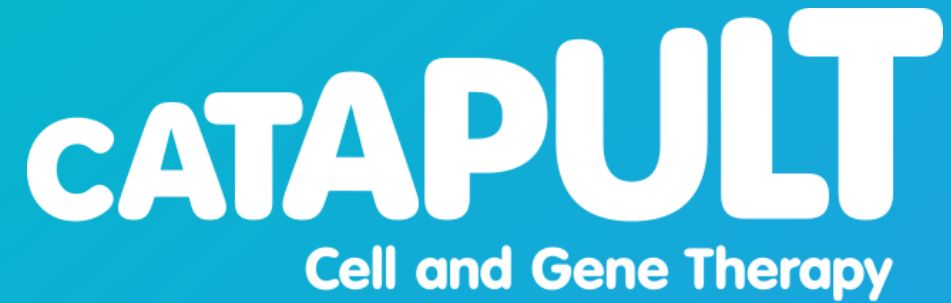
Imperial College London

Dr Ines Ullmo
Prof Hugh Brady

Earlham Institute

Dr Wilfried Haerty





Delivering impact.

12th Floor Tower Wing
Guy's Hospital
Great Maze Pond
London SE1 9RT

info@ct.catapult.org.uk
ct.catapult.org.uk
LinkedIn: cgt-catapult



Cell and Gene Therapy Catapult is committed to ensuring high standards of research integrity and research best practice in the activities we carry out. We subscribe to the principles described in the UK concordat to support research integrity. Cell and Gene Therapy Catapult is a trading name of Cell Therapy Catapult Limited, registered in England and Wales under company number 07964711, with registered office at 12th Floor Tower Wing, Guy's Hospital, Great Maze Pond, London, SE1 9RT. VAT number 154 4214 33.