

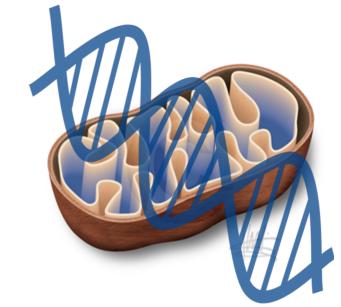
We assess the possibility of processing species monitoring surveys through eDNA with Oxford Nanopore MinION and a metagenomic sequencing approach. We use a mesocosom (fish tank where two live individuals of Alaskan chubs (Couesius plumbeus) for this study along with Halibut DNA as an internal control. Detection is possible despite a low sensitivity. Low variance of the detection rates is encouraging for further improvements.



Mitochondrial DNA for target:

- Macrobial eDNA in mitochondria and small cells (1),
- Target enlargement by >10 times,
- Abundance: in killifish, 0.4-2% of total nucleotide pool (2,3).





Oxford Nanopore Technology:

- Low-input barcoded metagenomic chemistry,
- minION: portability for field sequencing,
- real-time collection of data for

Monitoring surveys of elusive species:

- logistical efforts,
- financially costly,
- invasive for fragile species and ecosystems.



- **Environmental DNA (eDNA):**
- 2 methods: qPCR and metabarcoding,
- Species specific,
- extensive batteries of controls,
- obstacle for non-model organisms,
- Inaccessible to the public, \bullet
- Restricted from the field: analysis and response delays.

rapid-response actions.

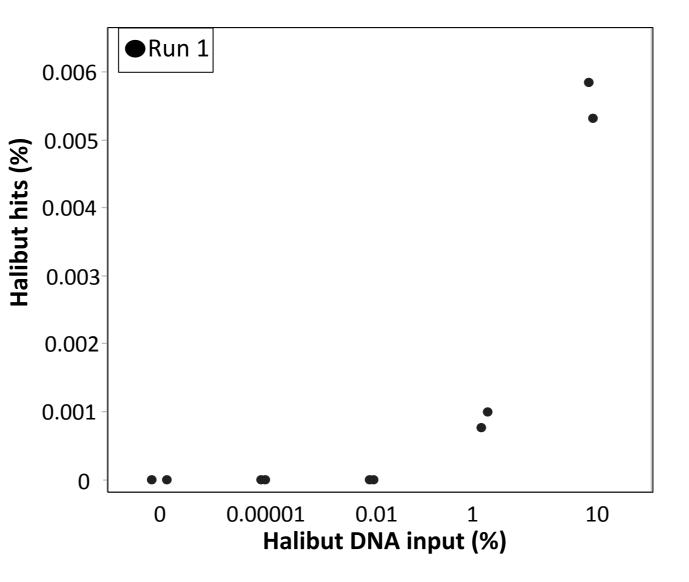
Perspectives:

- Development of a method optimized for detection limits,
- Assessment of success rate,
- Assessment of the optimized methodology in the field.

TESTING THE LIMITS OF DETECTION

eDNA extracted from fish tank water are spiked with Halibut DNA as an internal control. Experiment set in duplicate. Library prepared with the low-input barcoded kit, RPB004.

Halibut input	Barcode	yield (bp)	reads (n)	Halibut hits (n)	Couesius hits (n)
0%	2	696,841,147	234,680	0	1
	8	950,739,754	411,583	0	0
0.00001%	3	365,966,387	136,086	0	0
	9	757,419,597	270,119	0	1
0.01%	4	911,533,874	332,850	0	0
	10	439,116,212	151,480	0	1
1%	5	594,001,062	200,443	2	0
	11	668,766,201	260,830	2	0
10%	6	772,033,970	342,137	20	0
	12a	894,976,379	300,873	16	1
	Total:	7,051,394,583	2,641,081		



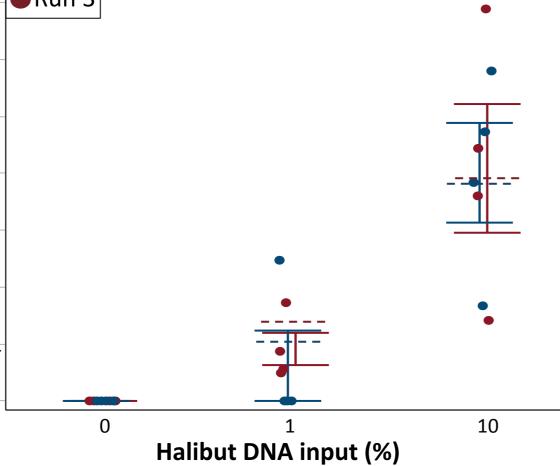
- Control spiking detected down to 1%: 50 pg or 16 pg/ul Halibut,
- Halibut detection correlates with internal control spiking,
- Duplicates show a low variance of detection,
- Fish species (*Couesius plumbeus*) from the fish tank is detected. \bullet

TESTING THE REPRODUCIBILITY

A second library is prepared with barcodes spiked within the limits of detection. Each spike is set in 4 replicates. The library is ran on 2 flow cells to assess the effect of sequencing depth on the reproducibility of detection.

Halibut input	Barcode	yield (bp)	reads (n)	Halibut hits (n)	Couesius hits (n)	0.007-	Run 2Run 3	
0%	9	195,717,621	62,486	0	0			
	10	141,536,046	48,095	0	1	⊗ ^{0.006−}		
	11	158,574,203	53,551	0	0	S 0 001		
	12a	252,942,817	83,755	0	0	-200.0 g		
1%	1	148,497,797	52,736	0	0	و <u>1</u> 0.004		
	2	184,314,450	69,253	0	0	- Halibut - But 0.003-		
	3	131,608,208	49,463	0	0	<u>.</u> 0.003-		
	4	233,893,642	80,901	2	0			•
10%	5	180,042,916	63,444	3	0	0.002-		
	6	161,409,106	59,826	1	0			
	7	222,491,787	78,163	3	0	0.001		
	8	191,711,649	68,993	4	0	0		1
	Total:	2,202,740,242	770,666			0-	0	1

Halibut input	Barcode	yield (bp)	ld (bp) reads (n)		Couesius hits (n)
0%	9	512,272,712	155,771	0	0
	10	326,694,036	106,770	0	0
	11	343,723,890	111,868	0	0
	12a	475,728,832	151,780	0	0
1%	1	331,566,073	114,696	1	0
	2	485,125,456	177,881	1	0
	3	317,761,543	115,847	2	0
	4	602,958,516	203,078	1	0
10%	5	414,557,260	141,332	2	1
	6	380,589,369	138,780	5	0
	7	553,885,865	188,746	13	1
	8	451,802,134	157,719	7	0
	Total:	5,196,665,686	1,764,268		



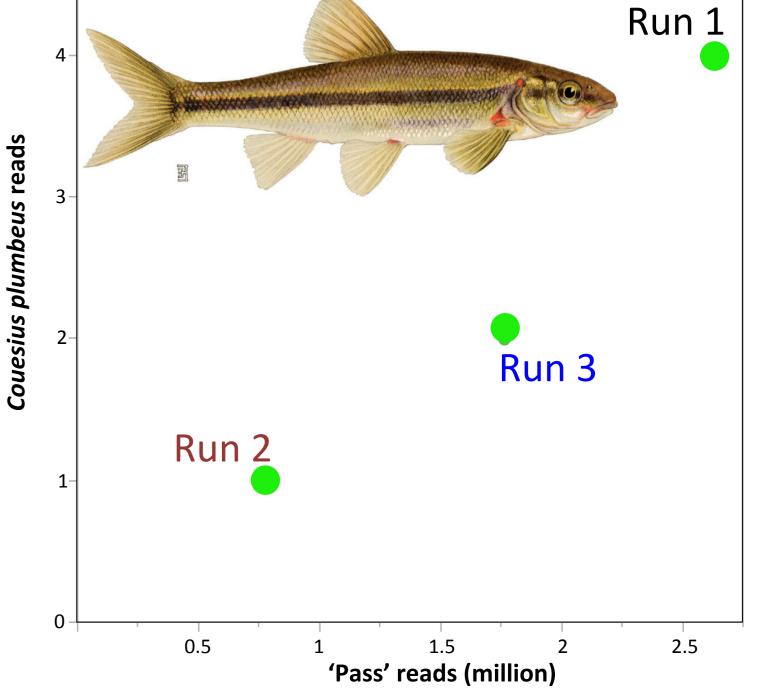
- Sequencing depth doesn't impact detection reproducibility,
- Low sensitivity of detection requires fair amounts of reads,
- 8 additional replicates of Halibut spiking consistent with run 1

DETECTION OF COUESIUS PLUMBEUS

Reads collected from the 3 runs are searched for mitochondrial hits of *Couesius plombeus*, the species of fish living in the fish tank.

• 2 individuals of fish living in 400L water detected with low-input metagenomic sequencing approach and mitochondrial hit search, **Detection success** correlates sequencing

depth.



<u>Controlled environment</u>: 400L fresh water fish tank (CFOS – UAF),

BIOINFORMATIC:

- Basecalling: ONT Albacore Sequencing Pipeline Software (version 2.2.7) - Quality filtering (qscore > 7)
- BLASTn (version 2.2.31+) search against custom mitochondrial database - word size 11 e-value cutoff 0.0001

References:

(1) Turner C. et al, 2014; Meth. in Ecol. and Evol.: 5 (7): 676-684. (2) Hartmann N. *et al*, 2011; *Aging Cell*: 10 (5): 824-831. (3) Valenzano D. et al, 2015; Cell: 163 (6): 1539-1554

Acknowledgements: We would like to thank the generous financial support of the BLaST program, the Institute of Arctic Biology, and Alaska INBRE. Research reported in this publication was supported by an Institutional Development Award (IDeA) from the National Institute of General Medical Sciences of the National Institutes of Health under grant number P20GM103395 as well as under three linked awards number RL5GM118990, TL4GM118992 and 1UL1GM118991. At UAF: J.A. López, P. Westley. From ONT: M. Micorescu and A..J. Markham.

- Living species: 2 Alaskan Lake chubs (*Couesius plumbeus;* mitochondrial genome NC_031568),
- <u>Sampling</u>: 24L of water; 2L per 0.45um Nitrocellulose filter,
 - DNA extraction: Qiagen PowerWater kit (lysis step modified with proteinase K),
 - DNA clean-up: MagBio magnetic beads,
 - DNA library preparation: Barcoded libraries RPB004 Rapid low-input barcoded kit (5ng input = 400 ml of sampled water), <u>Sequencing:</u> Oxford Nanopore minION; minKNOW 1.11.5; Flow cell R9.4.1